

Immunological laboratory testing in patients with tumours

The success of an “immunostimulatory” therapy for patients with tumour diseases essentially depends on the following points:

1. Is immunostimulation indicated?

- Immunostimulation must be indicated, i.e., it must be proven that those functional immune parameters are actually reduced. The stimulation of a functionally intact immune system is only helpful in exceptional cases. The current immunocompetence can only be evaluated through functional tests (LTT immune function, NK cell cytotoxicity test) and only with that it can successfully be controlled afterwards.
- It must be excluded that a pre-existing immune activation is not the cause of a reduced lymphocyte or NK cell function. In the cellular immune profile, therefore, the HLADR⁺ T cells should not be substantially elevated and the CD4/CD8 ratio should not be lower than 0.8.
- It must be proven that the patient has sufficient immune cells for stimulation (especially CD4⁺ helper cells). 250 CD4⁺ cells/μl are indicated as the limit. Otherwise, an immune-restorative therapy is preferable to immunostimulatory measures (trace elements, vitamins, thymus).

2. Is the immune system moving in the right direction during therapy?

During immunostimulatory therapy (e.g. with preparations containing mistletoe, organs or microorganisms), the following laboratory parameters can be consulted to evaluate progress and prognosis. Following questions can be addressed:

- **Was a TH1 polarisation achieved?**

The analysis is performed in the TH1/TH2 profile with measurement of Interferon-gamma (IFN-γ = TH1) and Interleukin-4 (IL-4 = TH2) after stimulation of the patient’s T lymphocytes. Basal T cell cytokine levels (in the blood) are not informative, because even in severe imbalances these cytokines IFN-γ / IL-4 are released into the blood in only insufficient quantities. The goal of immunostimulatory therapy is always a TH1 polarisation (i.e. IFN-γ ↑, IL-4 ↓). The IFN-γ-producing TH1 cells are the most important effector cells that contribute to eliminating “abnormal” cells.

IMD Labor Berlin		medical report	
Test	result	unit	reference range
TH1/TH2 - balance			
The cytokine concentrations after 24 hours are indicated. Stimulation with ConA/SEB.			
IFN-γ (TH1)	463	pg/ml	374 - 1660
IL-4 (TH2)	839	pg/ml	40 - 198
TH1 / TH2 ratio	0,55		3,5 - 11
The result shows a clear TH1/TH2 imbalance associated with the TH2 immune response.			

Fig. 1 Unfavourable TH2 predominance in a patient with metastatic rectal cancer before therapy. The goal of immunostimulatory therapy is an increase of IFN-γ with a decrease in IL-4.

- **Does the proportion of immunosuppressive regulatory T cells (Treg) remain low?**

The regulatory T cells (T_{reg}) are an important sub-group of the CD4 cells (normal approx. 4-10 %). They play a central role in the maintenance of immunotolerance, which is counterproductive in tumour illnesses. Regulatory T cells inhibit the effector functions of cytotoxic T cells, NK cells and other immune cells against tumour antigens and thus support tumour growth. A direct correlation between the number of T_{reg} cells and the stage of the tumour could be detected. Advanced tumour stages showed an increased infiltration of the tissue with Tregs. In addition, an inverse correlation between the number of T_{regs} in the tumour tissue and survival rate could be pointed out.

T_{reg} cells are well suited as progress markers in immunomodulating therapies. It is prognostically favourable if they do **not** rise. Regulatory T cells are phenotypically identified by flow cytometry based on the cell surface marker constellation CD4⁺CD25⁺⁺CD127^{low}.

In addition, the surface molecule CD39 is identified on T_{reg} cells. CD39 is a peripheral membrane protein (ectoenzyme), which causes the transformation of ATP and ADP into AMP. AMP is converted extracellularly into adenosine, which acts as an immunosuppressant. The detection of the CD39⁺ T_{reg} fraction can thus give a clue about the current immunosuppressive capacity of the T_{reg} cells.

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• **Do the CD8 cells with cytotoxic effector capacity increase?**

The term CD8 suppressor cells which is used incorrectly even today for the entire CD8 population is out-of-date. The "CD28 Status" differentiates the CD8 lymphocytes in cells with cytotoxic properties and those that actually perform a suppressive (immunosuppressive) function. As a matter of course the goal of immunostimulatory therapy is to increase the proportion of cytotoxic i.e. CD28-positive CD8 cells.

• **Do the activated natural killer cells increase?**

It is prognostically favourable, that during immunostimulatory therapy activation markers such as CD25 (receptor for interleukin-2) increase on natural killer cells. However, this test does not replace analysis of the NK cell function.

• **What is the patient's residual thymic function like?**

The surface marker CD31 defines a subpopulation of the naive CD4+ helper cells, which only recently left the thymus. These are referred to as "recent thymic emigrants – RTE cells". The proportion of the CD31+ naive CD4 cells in the blood is thus a measure of the residual thymic function. A decreased thymic function is exhibited in a reduced "ability to replenish" virgin T lymphocytes if there is an intensified "consumption" due to infections or after chemotherapy or radiotherapy. Therefore, the test is useful in the event of therapies that are immunologically damaging to assess the regeneration capacity and possible planning of early intervention (immune restoration marker).

The parameters mentioned are included in the quantitative immune profile "Immuno-competence Tumour" in addition to the recognised standard analyses such as CD4, CD8, NK and B cells and activated T cells.

Which diagnostic procedure is recommended?

Before immunostimulatory therapy

1. LTT immune function + NK cell cytotoxicity test (as an indication and a starting point)
2. Quantitative immune profile "Immuno-competence Tumour"
3. TH1/TH2 profile (as initial value)

The repetition of the analyses 6-8 weeks after beginning immunostimulation answers the following questions:

1. Did the function of the T lymphocytes and NK cells improve?
2. Is there a shifts in the cell markers which point toward a sustained improvement in the immunological situation (esp. T_{reg} cells and a portion of CD39+ Treg cells, CD28+ cytotoxic T cells)?
3. Could a TH1 polarisation be achieved (IFN-γ increase and/or IL-4 decrease)?

Subsequently, while check-ups 4-8 months apart are recommended regardless of the clinical situation, one may be limited to the pathological findings in the initial diagnosis.

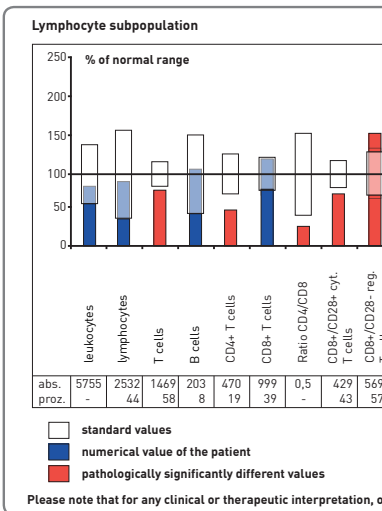


Fig. 2 The quantitative immune profile "Immuno-competence Tumour" was conducted as the initial state of a 58-year-old female patient with breast cancer. Due to the fact that regulatory T cells like CD8+/CD28+ as well as the percentage of CD39+ Tregs [CD4+/CD25+/CD127^{low}] are increased an immunostimulatory therapy should be carefully monitored. Regulatory cell subsets should not increase under immunostimulatory intervention.

The result is always displayed numerically and graphically.

		standard values		standard values	
Leukocytes	5755 /μl	3900 - 10200			
Lymphocytes	2532 /μl	1100 - 4500	44 %	20 - 44	
Monocytes	576 /μl	100 - 900	10 %	2 - 9,5	
Granulocytes	2647 /μl	2400 - 7400	46 %	42 - 75	
Immune competence					
T cells	1469 /μl	920 - 2580	58 %	61 - 84	
B cells	203 /μl	120 - 630	8 %	7 - 21	
NK cells	861 /μl	210 - 740	34 %	10 - 30	
CD4+ T helper cells	470 /μl	550 - 1460	19 %	32 - 60	
CD8+ T cells	999 /μl	280 - 930	39 %	23 - 40	
naive T cells [CD45RA+]	279 /μl	300 - 1200	19 %	30 - 63	
Thymic function [CD31+]			88 %	> 49	
Immune activation					
CD4/CD8 ratio	0,47	1 - 3			
activated T cells [HLA-DR+]	206 /μl	< 230	8 %	< 11	
pre-activated T cells [CD25+]	177 /μl	< 230	7 %	< 18	
memory T cells [CD45RO+]	1190 /μl	300 - 1300	81 %	37 - 70	
CD8+/CD28+ [cytotoxic]	429 /μl	238 - 448	43 %	49 - 73	
activated NK cells	23 /μl	< 40	2,7 %	< 17	
CD4+/CD8+ T cells			0,7 %	< 5	
Immune tolerance					
Treg [CD4+/CD25+/CD127 ^{low}]	22 /μl	35 - 120	4,7 %	4 - 10	
subset CD39+ Treg			64 %	< 54	
CD8+/CD28- (regulatory)	569 /μl	100 - 370	57 %	26 - 51	
CD8/CD28 ratio	0,75 /μl	1 - 2,8			
Immuno-competence: Intact – only slightly reduced CD4 cells					
Thymus reserve: Within the age-specific reference range					
Immune activation: Moderate signs – slightly elevated CD8- and NK- cells as well as an increased proportion of memory T cells [indication of chronic immune activation]					
Immune tolerance: Tendency towards an increase – increased regulatory [CD8+/CD28-] T cells and increased proportion of CD39+ Tregs [CD4+/CD25+/CD127 ^{low}], which may indicate functional immunosuppression.					

Explanatory representation of a very good progress finding after alternating therapy with mistletoe and a thymic preparation for a 54-year-old female patient with breast cancer.

1. LTT Immune Function

Preliminary findings

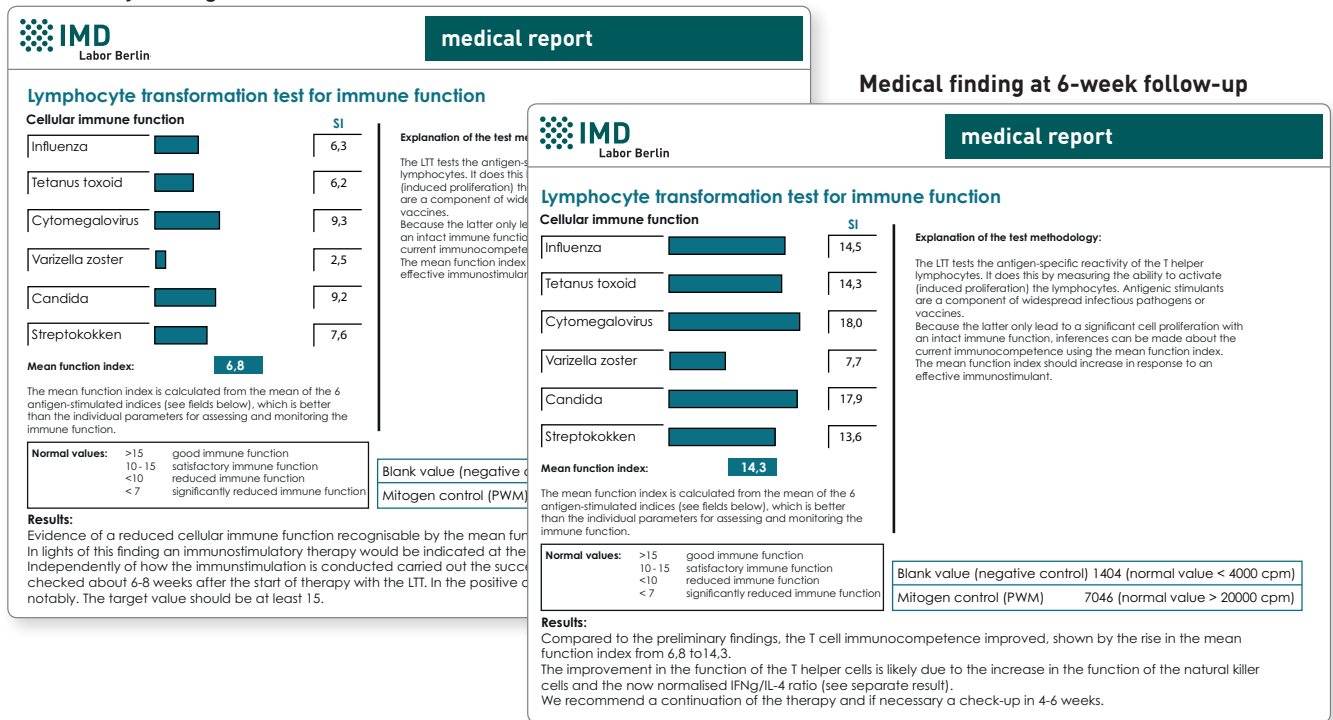


Fig. 3 The LTT immune function permits the analysis of the function of lymphocytes as well as indirectly monocytes and dendritic cells. The test is based on the principle of lymphocyte stimulation by memory antigens (recall antigens), against which should induce a strong immune response in intact cellular immunocompetence. When the assay is performed, these antigens are presented by monocytes and dendritic cells to T helper cells. These are activated in correlation with the current immune function and undergo cell division. The obligatory DNA synthesis is quantitatively detected.

2. NK cell cytotoxicity test and TH1/TH2-Profile

Preliminary findings

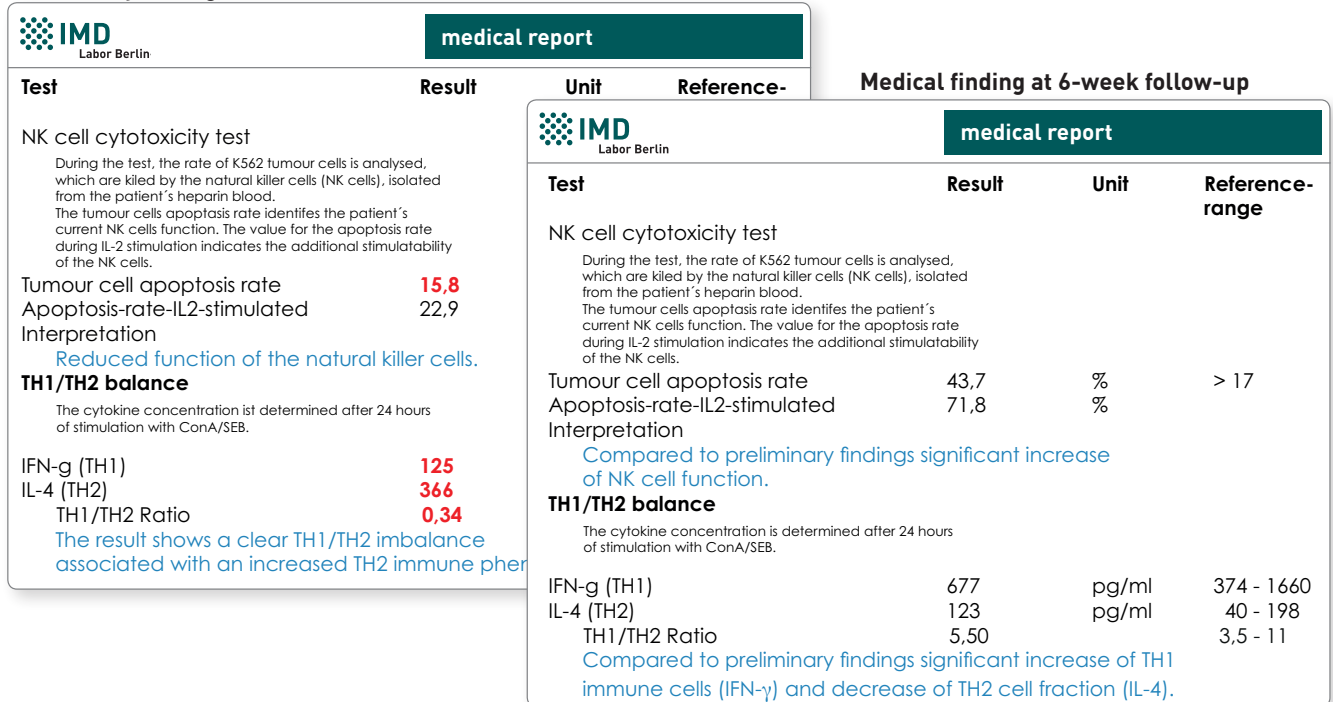


Fig. 4 For the NK cell cytotoxicity test, a tumour cell line is labelled with the membrane dye Calcein. Subsequently, the patient's NK cells are added under standardised conditions. During the lysis process induced by the NK cells, the Calcein dye is released from the tumour cells and then quantitatively determined in cell culture supernatant. By parallel testing of standardised controls, the percentage of destroyed tumour cells can be determined. To examine the TH1/TH2 balance heparinised blood is incubated with non-specific stimulants (Concanavalin A and SEB) for 24 h. Subsequently, the cytokines released into the supernatant are determined by multiplex cytokine assay.

3. Immune profile "Immunocompetence Tumour"

Preliminary findings

IMD Labor Berlin		medical report			
		standard values		standard values	
Leukocytes	4620 /µl	3900 - 10200			
Lymphocytes	1201 /µl	1100 - 4500	26 %	20 - 44	
Monocytes	416 /µl	100 - 900	9 %	2 - 9,5	
Granulocytes	3003 /µl	2400 - 7400	65 %	42 - 75	
Immune competence					
T cells	505 /µl	920 - 2580	42 %	61 - 84	
B cells	264 /µl	120 - 630	22 %	7 - 21	
NK cells	408 /µl	210 - 740	34 %	10 - 30	
CD4+ T helper cells	267 /µl	550 - 1460	22 %	32 - 60	
CD8+ T cells	227 /µl	280 - 930	19 %	23 - 40	
naive T cells (CD45RA+)	96 /µl	300 - 1200	19 %	30 - 63	
Thymus function (CD31+)			88 %	> 54	
Immune activation					
CD4/CD8 ratio	1,18	1 - 3			
activated T cells (HLA-DR+)	96 /µl	< 230	8 %	< 11	
pre-activated T cells (CD25+)	84 /µl	< 230	7 %	< 18	
memory T cells (CD45RO+)	409 /µl	300 - 1300	81 %	37 - 70	
CD8+/CD28+ [cytotoxic]	98 /µl	238 - 448	43 %	49 - 73	
activated NK cells	11 /µl	< 40	2,7 %	< 17	
CD4+/CD8+ T cells			0,4 %	< 5	
Immune tolerance					
Treg (CD4+/CD25+//CD127low)	13 /µl	35 - 120	4,7 %	4 - 10	
subset CD39+ Treg			64 %	> 54	
CD8-/CD28- (regulatory)	129 /µl	100 - 370	57 %	26 - 51	
CD8/CD28 ratio	0,75 /µl	1 - 2,8			

Immunocompetence: Reduced – CD4 and CD8 cells are reduced
Thymus reserve: Within the age-specific reference range
Immune activation: Subtle indications – increased proportion of memory T cells (indication of chronic immune activation)
Immune tolerance: Tendency towards increased – regulatory (CD8-/CD28-) T cells increased as well as a higher proportion of CD39+ Treg (CD4+/CD25+//CD127low) which may indicate functional immunosuppression.

Medical finding at 6-week follow-up

IMD Labor Berlin		medical report			
		standard values		standard values	
Leukocytes	5800 /µl	3900 - 10200			
Lymphocytes	1160 /µl	1100 - 4500	20 %	20 - 44	
Monocytes	348 /µl	100 - 900	6 %	2 - 9,5	
Granulocytes	4292 /µl	2400 - 7400	74 %	42 - 75	
Immune competence					
T cells	789 /µl	920 - 2580	68 %	61 - 84	
B cells	151 /µl	120 - 630	13 %	7 - 21	
NK cells	220 /µl	210 - 740	19 %	10 - 30	
CD4+ T helper cells	497 /µl	550 - 1460	43 %	32 - 60	
CD8+ T cells	260 /µl	280 - 930	22 %	23 - 40	
naive T cells (CD45RA+)	245 /µl	300 - 1200	31 %	30 - 63	
Thymus function (CD31+)			79 %	> 54	
Immune activation					
CD4/CD8 ratio	1,91	1 - 3			
activated T cells (HLA-DR+)	104 /µl	< 230	9 %	< 11	
pre-activated T cells (CD25+)	151 /µl	< 230	13 %	< 18	
memory T cells (CD45RO+)	544 /µl	300 - 1300	69 %	37 - 70	
CD8+/CD28+ [cytotoxic]	164 /µl	238 - 448	63 %	49 - 73	
activated NK cells	6 /µl	< 40	2,7 %	< 17	
CD4+/CD8+ T cells			0,1 %	< 5	
Immune tolerance					
Treg (CD4+/CD25+//CD127low)	27 /µl	35 - 120	5,4 %	4 - 10	
subset CD39+ Treg			51 %	> 54	
CD8-/CD28- (regulatory)	96 /µl	100 - 370	37 %	26 - 51	
CD8/CD28 ratio	1,7 /µl	1 - 2,8			

Compared to the preliminary finding, a tendency towards an increase in CD4 cells is observed. The fraction of cytotoxic (CD8+/CD28+) T cells has increased. Drop in the CD39+ Tregs with an almost unchanged overall proportion of CD4+/CD25+//CD127low Treg cells as well as a decline in the number of regulatory (CD8-/CD28-) T cells. In light of this finding, there is no contraindication to continue therapy.

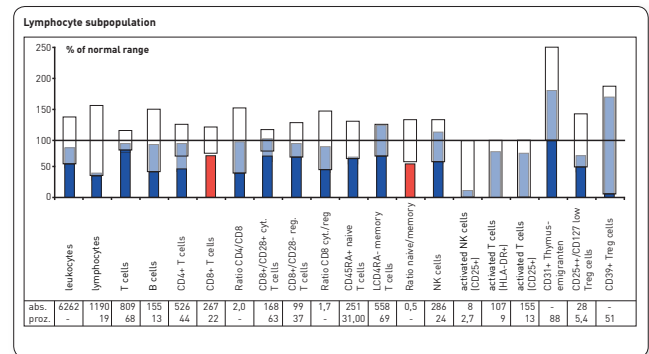
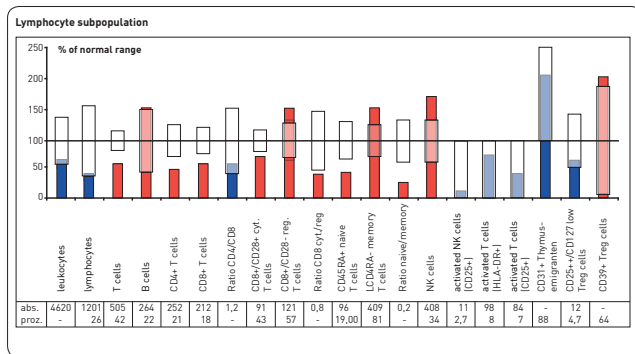


Fig. 5 The immune cells are differentiated based on the binding of fluorescence-marked monoclonal cell-specific antibodies and subsequent flow cytometry analysis. The graphical representation serves to better recognise deviations in cell distributions.

Material

LTT Immune Function:

20 ml heparin blood + 5 ml whole blood (serum)

NK cell cytotoxicity test:

10 ml heparin blood

Quantitative immune profile "Immunocompetence Tumour"

2 ml EDTA blood

TH1/TH2-Profile:

5 ml heparin blood

Costs

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Literature

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